

ab83463

Thioredoxin Reductase (TrxR) Assay Kit

Instructions for Use

For the rapid, sensitive and accurate measurement of Thioredoxin Reductase activity in various samples

This product is for research use only and is not intended for diagnostic use.

PLEASE NOTE: With the acquisition of BioVision by Abcam, we have made some changes to component names and packaging to better align with our global standards as we work towards environmental-friendly and efficient growth. You are receiving the same high-quality products as always, with no changes to specifications or protocols.

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1. Overview

Thioredoxin reductase (TrxR) (EC 1.8.1.9) is a ubiquitous enzyme which is involved in many cellular processes such as cell growth, p53 activity, and protection against oxidation stress, etc. The mammalian TrxR reduces thioredoxins as well as non-disulfide substrates such as selenite, lipoic acids, lipid hydroperoxides, and hydrogen peroxide.

Abcam's Thioredoxin Reductase (TrxR) Assay Kit provides a convenient colorimetric assay for detecting TrxR activity in various samples. In the assay, TrxR catalyzes the reduction of 5,5'-dithiobis (2-nitrobenzoic) acid (DTNB) with NADPH I/NADPH to 5-thio-2-nitrobenzoic acid (TNB²-), which generates a strong yellow color (λ_{max} = 412 nm). Since in crude biological samples other enzymes, such as glutathione reductase and glutathione peroxidase, can also reduce DTNB, therefore, TrxR specific inhibitor is utilized to determine TrxR specific activity.

Two assays are performed: the first measurement is of the total DTNB reduction by the sample, and the second one is the DTNB reduction by the sample in the presence of the TrxR specific inhibitor. The difference between the two results is the DTNB reduction by TrxR.

2. Protocol Summary

Sample Preparation

Standard Curve Preparation

Prepare and Add Reaction Mix

Measure Optical Density

3. Components and Storage

A. Kit Components

Item	Quantity
Thioredoxin Reductase Assay Buffer/TrxR Assay Buffer	25 mL
TNB Standard (Lyophilized)	1 vial
DTNB (Lyophilized)	1 vial
NADPH I/NADPH (Lyophilized)	1 vial
Thioredoxin Reductase Positive Control/TrxR Positive Control (Lyophilized)	1 vial
Thioredoxin Reductase Inhibitor/TrxR Inhibitor (Lyophilized)	1 vial

^{*} Store kit at -20°C, protect from light. Warm Thioredoxin Reductase Assay Buffer/Assay Buffer to room temperature before use. Briefly centrifuge all small vials prior to opening. Keep samples, NADPH I/NADPH, Thioredoxin Reductase Inhibitor/TrxR inhibitor, Thioredoxin Reductase Positive Control/TrxR Positive Control on ice during the assay. Read the entire protocol before performing the assay.

TNB STANDARD: Dissolve TNB Standard into 0.5 mL Thioredoxin Reductase Assay Buffer/Assay Buffer to generate 5 mM TNB

Standard. The TNB standard solution is stable for 1 week at +4°C or 2 month at -20°C.

DTNB SOLUTION: Dissolve DTNB into 0.9 mL Thioredoxin Reductase Assay Buffer/Assay Buffer, sufficient for 100 assays. The DTNB solution is stable for 1 week at +4°C or 2 month at -20°C.

NADPH I/NADPH: Dissolve one vial with 0.22 mL dH $_2$ O; sufficient for 100 assays. The solution is stable for 1 week at +4 $^{\circ}$ C or 2 month at -20 $^{\circ}$ C.

Thioredoxin Reductase Positive Control/TrxR POSITIVE CONTROL: Reconstitute with 90 µL Thioredoxin Reductase Assay Buffer/Assay Buffer to generate ~0.2 mU/µL TrxR; it is stable for 1 day at +4°C or 2 months at -20°C

Thioredoxin Reductase Inhibitor/TrxR INHIBITOR: Dissolve Thioredoxin Reductase Inhibitor/TrxR Inhibitor into 1.2 mL Thioredoxin Reductase Assay Buffer/Assay Buffer, sufficient for 100 assays. The Thioredoxin Reductase Inhibitor/TrxR Inhibitor solution is stable for 2 months at -20°C.

B. Additional Materials Required

- Microcentrifuge
- Bradford Reagent (ab102535)
- Protease Inhibitor Cocktail (ab65621)

- Pipettes and pipette tips
- Colorimetric microplate reader
- 96-well plate
- Orbital shaker

4. Assay Protocol

1. Sample Preparation:

Take 20 mg tissue or 2 x 10^6 cells and homogenize in 100- $200 \,\mu$ L cold Thioredoxin Reductase Assay Buffer/Assay Buffer on ice. Centrifuge at $10,000 \, x$ g for 15 min at +4°C; Collect the supernatant for assay and store on ice. Serum can be tested directly.

Determine the protein concentration of the cell or tissue supernatant using the Bradford Reagent (**ab102535**). Keep samples at -80°C for storage.

Note:

It is recommended to add Protease Inhibitor Cocktail (ab65621) to the buffer.

Add 2-50 μ L sample or 10 μ L Thioredoxin Reductase Positive Control/TrxR positive control into each well, adjusting volume to 50 μ L with Thioredoxin Reductase Assay Buffer/assay buffer.

2 sets of samples should be tested with or without Thioredoxin Reductase Inhibitor/TrxR Inhibitor. Add 10 μ L of Thioredoxin Reductase Inhibitor/TrxR Inhibitor to one set of the sample for testing background enzyme activity, and add 10 μ L of Thioredoxin Reductase Assay Buffer/Assay Buffer to the other set of sample for testing total DTNB reduction, mix well.

2. TNB Standard Curve:

Add 0, 2, 4, 6, 8, 10 μ L of the TNB Standard into 96-well plate in duplicate to generate 0, 10, 20, 30, 40, 50 nmol/well standard. Bring the final volume to 100 μ L with Thioredoxin Reductase Assay Buffer/Assay Buffer.

3. Reaction Mix:

Mix enough reagents for the number of assays to be performed. For each well, prepare a total 40 μ L Reaction Mix:

Thioredoxin Reductase Assay Buffer/Assay Buffer 30 µL

DTNB Solution 8 µL

NADPH I/NADPH 2 μL

Add 40 μL of the Reaction Mix to each test sample and positive control, mix well.

4. Measure OD_{412nm} at T_1 to get A_{1t} and A_{1l} , measure OD_{412nm} again at T_2 after incubating the reaction at 25°C for 20 min, to get A_{2t} and A_{2l} , protect from light. The incubation times can vary depending on the sample concentration.

The OD of TNB²⁻ generated by TrxR is:

$$\Delta A_{412nm} = (A_{2AB} - A_{2INH}) - (A_{1AB} - A_{1INH})$$

where AB is Thioredoxin Reductase Assay Buffer/assay buffer, INH is inhibitor

Note:

It is essential to read A1t, A1I, A2t and A2I' in the reaction linear range. It will be more accurate if you read the reaction kinetics. Then choose A1t, A1I, A2t and A2I in the reaction linear range.

Data Analysis

Plot the TNB standard curve. Apply the ΔA_{412nm} to the TNB standard curve to get ΔB nmol of TNB.

$$\frac{\text{TrxR}}{\text{Activity}} = \frac{\Delta B}{(T_2 - T_1) \times V} \times \text{Sample Dilution Factor} = \text{nmol/min/ml} = \text{mU/ml}$$

Where:

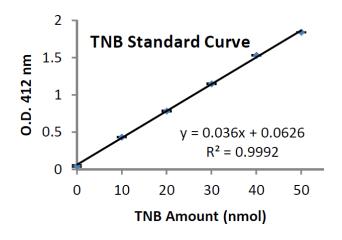
ΔB is the TNB amount from TNB standard curve (in nmol).

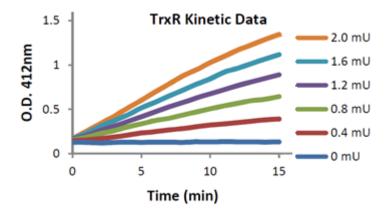
 T_1 is the time of the first reading $(A_{1t}, \text{ and } A_{1l})$ (in min).

 T_2 is the time of the second reading (A_{2t} and A_{2l}) (in min).

V is the pretreated sample volume (ml) added into the reaction well

Unit Definition: One unit of TrxR is the amount of enzyme that generates 1.0 μ mol of TNB per minute at 25°C. The oxidation of 1 mol of NADPH I/NADPH to NADP will generate 2 mol TNB finally, therefore, 1 TNB unit equals 0.5 NADP unit.





5. Troubleshooting

Problem	Reason	Solution	
Assay not working	Thioredoxin Reductase Assay Buffer/Assay buffer at wrong temperature	Thioredoxin Reductase Assay Buffer/Assay buffer must not be chilled - needs to be at RT	
	Protocol step missed	Re-read and follow the protocol exactly	
	Plate read at incorrect wavelength	Ensure you are using appropriate reader and filter settings (refer to datasheet)	
	Unsuitable microtiter plate for assay	Fluorescence: Black plates (clear bottoms); Luminescence: White plates; Colorimetry: Clear plates. If critical, datasheet will indicate whether to use flat- or U-shaped wells	
Unexpected results	Measured at wrong wavelength	Use appropriate reader and filter settings described in datasheet	
	Samples contain impeding substances	Troubleshoot and also consider deproteinizing samples	
	Unsuitable sample type	Use recommended samples types as listed on the datasheet	
	Sample readings are outside linear range	Concentrate/ dilute samples to be in linear range	

Samples with	Unsuitable sample type	Refer to datasheet for details about incompatible samples	
inconsistent readings	Samples prepared in the wrong buffer	Use the Thioredoxin Reductase Assay Buffer/assay buffer provided (or refer to datasheet for instructions)	
	Cell/ tissue samples not sufficiently homogenized	Increase sonication time/ number of strokes with the Dounce homogenizer	
	Too many freeze- thaw cycles	Aliquot samples to reduce the number of freeze-thaw cycles	
	Samples contain impeding substances	Troubleshoot and also consider deproteinizing samples	
	Samples are too old or incorrectly stored	Use freshly made samples and store at recommended temperature until use	
Lower/ Higher readings in	Not fully thawed kit components	Wait for components to thaw completely and gently mix prior use	
samples and standards	Out-of-date kit or incorrectly stored reagents	Always check expiry date and store kit components as recommended on the datasheet	
	Reagents sitting for extended periods on ice	Try to prepare a fresh reaction mix prior to each use	
	Incorrect incubation time/ temperature	Refer to datasheet for recommended incubation time and/ or temperature	
	Incorrect amounts used	Check pipette is calibrated correctly (always use smallest volume pipette that can pipette entire volume)	

Problem	Reason	Solution
Standard curve is not linear	Not fully thawed kit components	Wait for components to thaw completely and gently mix prior use
	Pipetting errors when setting up the standard curve	Try not to pipette too small volumes
	Incorrect pipetting when preparing the reaction mix	Always prepare a master mix
	Air bubbles in wells	Air bubbles will interfere with readings; try to avoid producing air bubbles and always remove bubbles prior to reading plates
	Concentration of standard stock incorrect	Recheck datasheet for recommended concentrations of standard stocks
	Errors in standard curve calculations	Refer to datasheet and re-check the calculations
	Use of other reagents than those provided with the kit	Use fresh components from the same kit



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